

# **COMMENTARY**

# Selective block of KATP channels: why the anti-diabetic sulphonylureas and rosiglitazone have more in common than we thought

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Rosiglitazone, the thiazolidinedione class anti-diabetic withdrawn from Europe in 2010 amid reports of adverse cardiovascular effects, is revealed by Yu et al. in this issue of the British Journal of Pharmacology to be a selective blocker of ATP-sensitive potassium (K<sub>ATP</sub>) channels. This seems little cause for excitement given that the closure of pancreatic K<sub>ATP</sub> channels is integral to insulin secretion; and sulphonylureas, which inhibit  $K_{ATP}$  channels, are widely used to treat type II diabetes. However, rosiglitazone, whose primary targets are nuclear transcription factors that regulate genes involved in lipid metabolism, blocks K<sub>ATP</sub> channels by a novel mechanism different to that of the sulphonylureas and has a worrying preference for blood flow-regulating vascular K<sub>ATP</sub> channels. Identification of a new molecule that modulates K<sub>ATP</sub> channel gating will not only tell us more about how these complex metabolic sensors work but also raises questions as to whether rosiglitazone suppresses the cardiovascular system's ability to cope with metabolic stress – a claim that has dogged the sulphonylureas for many years.

#### LINKED ARTICLE

This article is a commentary on Yu et al., pp. 26-36 of this issue. To view this paper visit http://dx.doi.org/10.1111/ j.1476-5381.2012.01934.x

#### **Abbreviations**

ER, endoplasmic reticulum; K<sub>ATP</sub> channel, ATP-sensitive potassium channel; SUR, sulphonylurea receptor

The anti-diabetic rosiglitazone has had a chequered history. Working as an agonist at nuclear PPARy, it was introduced in 1999 and widely prescribed for the treatment of type II diabetes mellitus based on its ability to increase insulin sensitivity in fat cells by regulating genes involved in glucose and lipid metabolism (Brown and Plutzky, 2007). Additional beneficial cardiovascular effects were soon linked to its usage, including anti-inflammatory and anti-proliferative effects that retarded the development of atherosclerosis (Zinn et al., 2008). These wholly positive outcomes were tempered by reports that rosiglitazone and its fellow thiazolidinediones exacerbated fluid retention and congestive heart failure (Zinn et al., 2008; Kaul et al., 2010), although the drug's benefits were still considered to largely outweigh any risks. Opinion shifted in 2007 with the publication in the New England Journal of Medicine of a large-scale meta-analysis of 42 randomized trials involving almost 28 000 patients that indicated an alarming 43% increase in myocardial infarction in

patients taking rosiglitazone (Nissen and Wolski, 2007). Publication of this report prompted the United States Food and Drug Administration (FDA) to release a safety alert flagging the possible increased risk of ischaemic cardiovascular events in patients prescribed rosiglitazone. The ensuing controversy saw a flurry of additional publications supporting or refuting the adverse cardiovascular effects of the drug (reviewed by Zinn et al., 2008; Kaul et al., 2010), muddying the waters to such an extent that the FDA Advisory Panel subsequently voted against removing rosiglitazone from the US market. The European Medicine Agency took a harder line and withdrew rosiglitazone from Europe in September 2010. Whether rosiglitazone produces net clinical benefit or harm is still far from clear, and in this issue of the British Journal of Pharmacology, Yu et al. add to the debate by revealing that rosiglitazone at near clinically relevant concentrations acts to inhibit ATP-sensitive potassium (K<sub>ATP</sub>) channels, a family of proteins that play critical protective roles during acute metabolic stress



(Yu et al., 2012). While rosiglitazone-induced block of KATP channels is not in itself novel, Yu et al. showed that unlike the well-characterized sulphonylureas that inhibit K<sub>ATP</sub> channels by interacting with their large modulatory sulphonylurea receptor (SUR) subunit, rosiglitazone suppresses the channel's open probability by interacting with the cytosolic face of pore-forming K<sub>IR</sub>6.x subunit. They also demonstrated a 4-fold increase in rosiglitazone potency for K<sub>ATP</sub> channels containing the K<sub>IR</sub>6.1 pore-forming subtype. Given the primarily vascular distribution of K<sub>IR</sub>6.1-containing channels, this raises a number of questions with regard to the potential clinical implications of rosiglitazone usage. This also echoes the ongoing debate regarding the cardiovascular safety of another mainstay of type II diabetes, the sulphonylureas (Tzoulaki et al., 2009).

An anti-diabetic drug that selectively inhibits K<sub>ATP</sub> channels seems on the face of it to be little cause for concern. Inhibition of these channels by high ATP (glucose) levels in pancreatic beta cells induces membrane depolarization, Ca<sup>2+</sup> influx via voltage-gated Ca2+ channels and Ca2+-dependent secretion of insulin (Ashcroft and Gribble, 1999). Indeed, sulphonylureas such as tolbutamide, glibenclamide (glyburide) and glimepiride have been used clinically to treat type II diabetes mellitus for many years, often in combination with the insulin-sensitizing thiazolidinediones. Pancreatic betacell K<sub>ATP</sub> channels most likely form as octomers of four poreforming  $K_{IR}6.2$  and 4 modulatory SUR1 subunits, but other distinct K<sub>ATP</sub> channel isoforms also exist in cardiac and smooth muscle. Activation of vascular K<sub>ATP</sub> channels (K<sub>IR</sub>6.1/ SUR2B) by vasodilating transmitters causes membrane hyperpolarization, decreased Ca2+ entry and vasorelaxation (Flagg et al., 2010), and drugs that open vascular KATP channels to increase arterial diameter and blood flow are used to treat angina pectoris (nicorandil) and intractable hypertension (minoxidil and diazoxide). The cardiac isoform of  $K_{\text{ATP}}$ channel (K<sub>IR</sub>6.2/SUR2A) opens during ischaemia, promoting membrane repolarization and a shortening of the action potential, which reduces Ca<sup>2+</sup> entry in an attempt to conserve ATP and thus minimize cell damage (Flagg et al., 2010). Blockade of vascular K<sub>ATP</sub> channels by rosiglitazone would therefore be expected to have adverse effects at times when the coronary circulation needs to dilate, for example during exercise or stress, and inhibition of cardiac K<sub>ATP</sub> channels has been shown to severely compromise the heart's ability to cope with ischaemic assault. What the study by Yu et al. highlights is that micromolar concentrations of rosiglitazone inhibit the activity of all isoforms of KATP channel, but with a marked preference for  $K_{ATP}$  channels containing the  $K_{IR}6.1$  isoform. The IC<sub>50</sub> of rosiglitazone is calculated to be 45  $\mu$ M for K<sub>IR</sub>6.2/ SURx (pancreatic and heart) channels and 10 µM for the vascular K<sub>IR</sub>6.1/SUR2B, which is reduced to 2 μM in the presence of therapeutic concentrations of sulphonylureas (Yu et al., 2011). Plasma concentrations of rosiglitazone in the treatment of type II diabetes are generally in the region of 3 μM (Cox et al., 2000), which places the vascular channel in particular in the firing line. A caveat to this is that the IC<sub>50</sub> values stated above were obtained by directly applying rosiglitazone to the cytoplasmic face of the channel in excised inside-out membrane patches. The effects of rosiglitazone were considerably weaker when applied extracellularly. This difference in potency was taken by the authors to indicate

that rosiglitazone acts primarily on the cytosolic face of the channel and that the weakened effects associated with extracellular application were due to the drug having to cross the membrane to reach its active site. It should be remembered that rosiglitazone's primary targets are nuclear transcription factors and the drug has the ability to enter cells quite readily. Indeed, the authors have previously shown that 30 µM rosiglitazone reduces by a third the ability of the coronary arteries to dilate in response to  $\beta$ -adrenoceptor stimulation in a Langendorff-perfused heart preparation (Yu et al., 2011). Thus, while it is unclear if clinical doses of rosiglitazone would reach levels sufficient to significantly inhibit vascular K<sub>ATP</sub> channels, the potential for rosiglitazone to uncouple the coronary circulation from autonomic control is serious enough to warrant further investigation. Similar unresolved fears have of course existed since the early 1970s regarding the sulphonylureas and their ability to block both cardiac and vascular K<sub>ATP</sub> channels.

Yu et al. use K<sub>IR</sub>6.2ΔC36 channels to show that rosiglitazone acts predominantly on the pore-forming K<sub>IR</sub>6.x subunit. These truncated subunits lack a C-terminal ER retention signal and form functional channels at the cell surface without the SUR subunit (Zerangue et al., 1999).  $K_{IR}6.2\Delta C36$  channels are as sensitive to rosiglitazone as the complete K<sub>IR</sub>6.2/SURx complex, which distinguishes rosiglitazone's action from that of the sulphonylureas and places it in the same class as the pore-acting K<sub>ATP</sub> channel blocker PNU-37883A. As discussed above, differences in the potency of rosiglitazone when applied to the intracellular or extracellular side of the membrane also led the authors to conclude that the main site of action is on the pore-forming subunit's intracellular domain. These intracellular regions contain the sites where ATP binds to inhibit channel activity and an amphipathic 'slide' helix that lies parallel to the cytosolic face of the membrane (Nichols, 2006). This helix is tethered to the cytoplasmic end of the channel pore where the channel 'gate' is believed to reside and thus may link ATP binding to opening/closure of the potassium conduction pathway. Analysis of the behaviour of single K<sub>ATP</sub> channels in the presence and absence of rosiglitazone shows that while the drug has no effect on the size of single channel openings, it suppresses channel activity by extending longlasting channel closures, almost certainly by modulating the channel's complex gating mechanism. Channel openings or closures result from the balance between the inhibitory drive of ATP binding to the pore-forming subunit and the activating drive of MgADP binding the modulatory SUR subunit. Additionally, membrane lipids such as phosphatidylinositol 4,5-bisphosphate interact with sites on K<sub>IR</sub>6.x that overlap with the ATP binding site and promote channel activity by antagonizing ATP inhibition (Flagg et al., 2010). Clues as to rosiglitazone's action may come from the fact that K<sub>IR</sub>6.1 and 6.2 share around 70% amino acid sequence identity, with much of the divergency occurring in the C-terminal intracellular region. Identification of rosiglitazone's exact binding site and mechanism of action may provide additional insight into the channel's gating mechanism and the reason why rosiglitazone has a fourfold preference for K<sub>IR</sub>6.1. This in turn may highlight key differences between the pore-forming isoforms that aid the design of isoform-specific modulators.

# Selective block of K<sub>ATP</sub> channels



Rosiglitazone is unlikely to be re-introduced into the European market anytime in the near future but is still available under restriction in the States. Despite many positive reports on its clinical cardiovascular effects (reviewed by Zinn et al., 2008), there remains the lingering suspicion that in certain patients under certain conditions, rosiglitazone promotes unfavourable outcomes. The finding that it acts as a selective inhibitor of KATP channel and may thus inadvertently inhibit a population of ion channels involved in ischaemic protection and the regulation of blood flow may help focus future clinical studies.

## References

Ashcroft FM, Gribble FM (1999). ATP-sensitive K+ channels and insulin secretion: their role in health and disease. Diabetologia 42: 903-919.

Brown JD, Plutzky J (2007). Peroxisome proliferator-activated receptors as transcriptional nodal points and therapeutic targets. Circulation 115: 518-533.

Cox PJ, Ryan DA, Hollis FJ, Harris AM, Miller AK, Vousden M et al. (2000). Absorption, disposition, and metabolism of rosiglitazone, a potent thiazolidinedione insulin sensitizer, in humans. Drug Metab Dispos 28: 772-780.

Flagg TP, Enkvetchakul D, Koster JC, Nichols CG (2010). Muscle K(ATP) channels: recent insights to energy sensing and myoprotection. Physiol Rev 90: 799-829.

Kaul S, Bolger AF, Herrington D, Giugliano RP, Eckel RH (2010). Thiazolidinedione drugs and cardiovascular risks a science advisory from the American Heart Association and American College of Cardiology Foundation. Circulation 121: 1868–1877.

Nichols CG (2006). K-ATP channels as molecular sensors of cellular metabolism. Nature 440: 470-476.

Nissen SE, Wolski K (2007). Effect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular causes. N Engl J Med 356: 2457-2471.

Tzoulaki I, Molokhia M, Curcin V, Little MP, Millett CJ, Ng A et al. (2009). Risk of cardiovascular disease and all cause mortality among patients with type 2 diabetes prescribed oral antidiabetes drugs: retrospective cohort study using UK general practice research database. Br Med J 339: b4731.

Yu L, Jin X, Yang Y, Cui NR, Jiang C (2011). Rosiglitazone inhibits vascular KATP channels and coronary vasodilation produced by isoprenaline. Br J Pharmacol 164: 2064-2072.

Yu L. Jin X. Cui NR. Wu Y. Shi Z. Zhu D et al. (2012). Rosiglitazone selectively inhibits KATP channels by acting on the Kir6 subunit. Br J Pharmacol 167: 26-36.

Zerangue N, Schwappach B, Jan YN, Jan LY (1999). A new ER trafficking signal regulates the subunit stoichiometry of plasma membrane K-ATP channels. Neuron 22: 537-548.

Zinn A, Felson S, Fisher E, Schwartzbard A (2008). Reassessing the cardiovascular risks and benefits of thiazolidinediones. Clin Cardiol 31: 397-403.